Full Length Research Paper

In vitro antiviral activity of diterpenes isolated from the Brazilian brown alga *Canistrocarpus cervicornis*

Magui Aparecida Vallim¹, Juliana Eymara Barbosa¹, Diana Negrão Cavalcanti¹, Joel Campos De-Paula², Viveca Antonia Giongo Galvão da Silva¹, Valéria Laneuville Teixeira^{1*} and Izabel Christina Nunes de Palmer Paixão^{1,3}

¹Programa de Pós-Graduação em Biologia Marinha, Instituto de Biologia, Universidade Federal Fluminense, P. O.Box 100.644, CEP 24001-970, Niterói, RJ, Brazil.

²Instituto de Biociências, Universidade Federal do Estado do Rio de Janeiro, Avenida Pasteur 458, CEP 22290-240, Urca, Rio de Janeiro, RJ, Brazil.

³Departamento de Biologia Celular e Molecular, Instituto de Biologia, Universidade Federal Fluminense, Niterói, RJ, Brazil.

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They describe in this paper that the dolastane diterpenes 4-hydroxy-9,14-dihydroxydolasta-1(15),7diene (1) and 4,7,14-trihydroxydolasta-1(15),8-diene (2), isolated from the marine brown alga *Canistrocarpus cervicornis* inhibited HSV-1 infection in Vero cells. The concentration required to inhibit HSV-1 replication was not cytotoxic. Their results suggest that the structures of diterpenes 1 and 2, dolastane diterpenes from Brazilian brown algae, are promising for future antiviral drugs.

Key words: *Canistrocarpus cervicornis*, dictyotaceae, phaeophyceae, dolastane diterpene, cytotoxicity, antiviral against HSV-1.

INTRODUCTION

Species of the family Dictyotaceae produce a large array of bioactive secondary metabolites and have a broad defensive action against herbivores in the marine environment. These species have been shown to exhibit antifungal, antibacterial (Zanfardino and Gavagnin, 2009) and antiviral activities (Abrantes et al., 2009; Barbosa et al., 2004; Cirne-Santos et al., 2006; 2008; Pereira et al., 2004; 2005). Phycochemical studies have been undertaken on the family Dictyotaceae resulting in the isolation of more than 300 diterpenes from at least 35 species collected all over the world (Vallim et al., 2005).

Canistrocarpus cervicornis (Kützing) De Paula and De Clerck (De Clerck et al., 2006) is an important and abundant alga from the Brazilian coast (Falcão and Menezes de Széchy 2005). Although, many diterpenes have been isolated from Brazilian *C. cervicornis* (e.g. Fleury et al., 1994; Kelecom and Teixeira 1988; Teixeira et al., 1986a, b), the antiviral activity of this type of compound is unknown. The Figure 1 presents the structure of dolastane diterpenes 1 and 2 and the previously reported the antiviral activity against HIV-1 (Human Immunodeficiency virus type 1) and HSV-1 (Herpes simplex virus type 1) of dolabellane (3-5) and dichotomane (6-7) diterpenes (Abrantes et al., 2009; Barbosa et al., 2004; Cirne-Santos et al., 2006; 2008; Pereira et al., 2004; 2005).

The HSV-1 infections may cause oral disorders, keratoconjutivitis and encephalitis infection that varies in severity from sub-clinical infections to fatal ones. Most of clinical anti-herpes virus compounds are nucleoside analogues, such as acyclovir (ACV), the most common drug used on the treatment of HSV infections (Brady and Bernstein, 2004). Nonetheless, drug-resistant HSV strains frequently emerge during a long ACV-based treatment due to the reduced expression or the not functioning of timidine kinase. Therefore, the search for new antiviral agents, especially with different mechanisms

^{*}Corresponding author. E-mail: valerialaneuville@gmail.com. Tel: +55 021 21 26292296. Fax: +55 021 21 2629 2292. 2380 J. Med. Plant. Res.

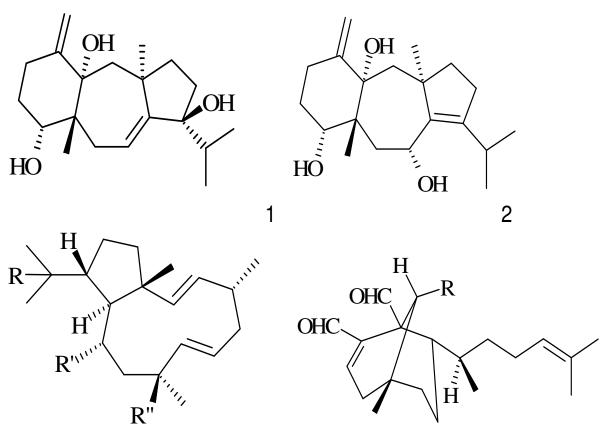


Figure 1. Dolastane diterpenes 1 and 2 isolated from Brazilian C. cervicornis.3 R = R' = OAc, R'' = OH6 R = OH4 R = R' = OH, R'' = OAc7 R = OAc5 R = R' = R'' = OH.

of actions, is a crucial goal (De Clerq, 2004; Morfin and Thouvenot, 2003; Newman et al., 2003). The present study, we evaluate for the first time the antiviral activity against HSV-1 of dolastane diterpenes isolated from species *C. cervicornis*.

MATERIALS AND METHODS

Specimens of C. cervicornis were collected at Búzios, Rio de Janeiro, Brazil, and they were identified by De-Paula, one of the authors of this paper. The seaweeds were washed with local sea water, separated from sediments, epiphytes and other associated organisms. Voucher specimens was deposited in the Herbarium of the Universidade do Estado do Rio de Janeiro (HRJ 10754). The diterpene 4-hydroxy-9,14-dihydroxydolasta-1(15),7-diene (1) was isolated from the CH₂Cl₂ crude extract of C. cervicornis after column chromatography on silica gel. The fractions eluted in CH₂Cl₂ 100% contained the dolastane diterpene 1. The fractions eluted with CH2Cl2/EtOAc 7:3 to CH2Cl2/EtOAc 1:1 yielded crude diterpene 4,7,14-trihydroxydolasta-1(15),8-diene (2). The combined fractions were subjected to silica gel column chromatography. The fractions eluted with n-hexane/ EtOAc (3:7) yielded the diterpene 2. The compounds 1 and 2 were identified by comparison of physical and spectroscopic data with reported values (Crews et al., 1982; González et al., 1983; Teixeira et al., 1986a). The diterpenes, at

over 99% purity, were diluted in 100% dimethyl sulfoxide (DMSO) and stored at -20°C. The resulting DMSO concentrations during the assays were below 0.1%, a level that is not significantly cytotoxic. African green monkeys kidneys cells (Vero cells; ATCC) were cultured in Dulbecco's modified Eagle's medium (DMEM; GIBCO) supplemented with 5% fetal bovine serum (FBS; HyClone), 100 U/mL of penicillin, 100 µg/mL of streptomycin and 2.5 µg / mL of amphotericin B (Fungizon) and incubated at 37°C in 5% CO₂. The Vero cells were treated with PBS-1X/EDTA (Reagen) and 0.25% trypsin (GIBCO) so as to achieve the experimental conditions. HSV-1 was kindly provided by Dra. Marcia Wigg (Universidade Federal do Rio de Janeiro, Brazil). Virus stocks were stored at -70°C until use. Monolayers of Vero cells in 96-multiwell plates (1 × 10⁴/well) were treated with several concentrations (50, 250, 500 and 1000 µM) of diterpenes 1 and 2 of the C. cervicornis for 72 h at 37°C in atmosphere of 5% CO2. Then, 50 µL of a 1 mg/mL solution of 3-4,5dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide (MTT; Sigma) was added to evaluate the cell viability according to procedures described elsewhere (Gilbert et al., 2002). The cytotoxic concentration in 50% (CC₅₀) was calculated by linear regression analysis of the dose-response curves generated from the data using Microsoft Excel. All experiments were performed in triplicate at least three times.

The antiviral activities of 1 and 2 were investigated according to Reed and Munch 50% end point method (Reed and Munch, 1938). At first, Vero cells grown to confluence in 96-well plate were infected with HSV-1 (AR-29) at 1 multiplicity of infection for 1 h at Vallim et al. 2381

Table 1. Dose-dependent cytotoxicity and percentage of inhibition of viral replication observed in Vero cells in the presence of the diterpenes 1 and 2 from the brown alga *C. cervicornis*.

Sample	СС ₅₀ (µМ)	% Inhibition ^(*)
Diterpene 1	1423 (± 83)	90
Diterpene 2	706 (± 100)	99
Acyclovir	860 (± 32)	99

(*) Base on TCID₅₀ which means could determine a median tissue culture infective dose that amount of a pathogenic agent that will produce pathological change in 50% of Vero cell cultures incubated with $50\mu M$ of the substances.

37°C. After that, infected cells were washed with PBS to remove residual viruses and a complete culture medium, with or without diterpenes, were added. 72 h after infection, the virus titer of each sample was determined in terms of the 50% tissue culture dose (TCID₅₀/mL) by end point dilution. In addition we used mock-treated HSV1-infected cells as positive control. To evaluate the virucidal activity of the diterpenes 1 and 2 from *C. cervicornis*, we conducted cell-free tests with viral suspension in a medium without FCS at a concentration of $10^7 - 10^5$ particles/mL in the presence and in the absence of the substances (50 μ M). The samples were incubated with viral suspension for periods of 1, 2 and 4 h at 4°C.

RESULTS AND DISCUSSION

We observed that both compounds were not cytotoxic for Vero cell lineage (Table 1). We also observed that the CC_{50} of the diterpene 1 was 48% higher than acyclovir's. Acyclovir is more selective for viral DNA polymerase than for cellular enzyme, having minimal toxicity (Brady and Bernstein, 2004). This finding is very promising for future antiviral trials. A reduction of 3 logs of infectivity was evident for diterpene 2 of C. cervicornis when compared to the values obtained with HSV-1 control (Table 1) according to TCID₅₀ method. Although, both compounds have shown viral inhibition rates above 90% we observed specifically for diterpene 2 the same inhibitory rate of acyclovir. For this reason, our future studies are concerned about the kinetic activity of polymerase from HSV-1 in the presence of these compounds. In the virucidal assay, we discovered that only after 4 h of incubation did the diterpene 2 show virucidal activity at the dilution of 10⁶ (78%) and for this reason we cannot say that these compounds may be presented as possible virucidal molecules (data not showed).

Many investigators have reported the inhibitory effects of algal extracts and their compounds on the replication of HSV-1, with special emphasis in sulfated polysaccharides (Damonte et al., 2004; Harden et al., 2009; Witvrouw and De Clercq, 1997). In this study, we showed the antiviral activity of diterpenes obtained from *C. cervicornis*. Both diterpenes 1 and 2 extracted from *C. cerviconis* were able to inhibit *in vitro* replication of herpes simplex virus type 1. However, the viral inhibition rate obtained with diterpene 2 was higher than that of diterpene 1. The presence of the 7-hydroxyl group can have a positive influence on the inhibition of HSV-1. Since the same profile of antiviral diterpenes could not be observed in the absence of cells, that is, these substances will not be considered virucidal. Thus, we considered these substances promising as anti-HSV-1 and therefore able to advance to the *in vivo* antiviral studies.

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REFERENCES

- Abrantes JL, Barbosa J, Cavalcanti D, Pereira RC, Fontes CFL, Teixeira VL, Souza TML, Paixão ICP (2009). The effects of the diterpenes isolated from the Brazilian brown algae *Dictyota pfaffii* and *Dictyota menstrualis* against the herpes simplex type-1 replicative cycle. Planta Med., 75: 1–6.
- Barbosa JP, Pereira RC, Abrantes JL, Cirne dos Santos CC, Rebello MA, Frugulhetti IC, Texeira VL (2004). *In vitro* antiviral diterpenes from the Brazilian brown alga *Dictyota pfaffii*. Planta Med., 70: 856–860.
- Brady RC, Bernstein DI (2004). Treatment of herpes simplex virus infections. Antiviral Res., 61: 73-81.
- Cirne-Santos CC, Teixeira VL, Castello-Branco LR, Frugulhetti ICPP, Bou-Habib DC (2006). Inhibition of HIV-1 replication in human primary cells by a dolabellane diterpene isolated from the marine algae *Dictyota pfaffii*. Planta Med., 72: 295-299.
- Cirne-Santos CC, Souza TML, Teixeira VL, Fontes CFL, Rebello MA, Castello-Branco LR, Abreu CM, Tanuri A, Frugulhetti ICPP, Bou-Habib DC (2008). The dolabellane diterpene Dolabelladienetriol is a typical noncompetitive inhibitor of HIV-1 reverse transcriptase enzyme. Antiviral Res., 77: 64-71.
- Crews P, Klein TE, Hogue ER, Myers BL (1982). Tricyclic diterpenes from the brown marine algae *Dictyota divaricata* and *Dictyota linearis*. J. Org. Chem., 47: 811-815.
- Damonte, EB, Matulewicz MC, Cerezo AS (2004). Sulfated seaweed polysaccharides as antiviral agents. Curr. Med. Chem., 11: 2399-2419.
- De Clercq E (2004). Antiviral drugs in current clinical use. J. Clin. Virol., 30: 115-133.

De Clerck O, Leliaert F, Verbruggen H, Lane CE, De-Paula JC, Payo DA, Coppejans E (2006). A revised classification of the Dictyoteae

2382 J. Med. Plant. Res.

- (Dictyotales, Phaeophyceae) based on rbc and 26S ribosomal DNA sequence analyses. J. Phycol., 42: 1271-1288.
- Falcão C, Menezes de Széchy MT (2005). Changes in shallow phytobenthic assemblages in southeastern Brazil following the replacement of *Sargassum vulgare* (Phaeophyta) by *Caulerpa scalpelliformis* (Chlorophyta). Bot. Mar., 48: 208-217.
- Fleury BG, Kelecom A, Pereira RC, Teixeira, VL (1994). Polyphenols terpenes and sterols in Brazilian Dictyotales and Fucales. Bot. Mar., 37: 457-462.
- Gilbert C, Bestman-Smith J, Boivin G (2002). Resistance of Herpesviruses to Antiviral Drugs: Clinical Impacts and Molecular Mechanisms. Drug Resist. Update, 5: 88-114.
- González AG, Martín JD, Norte M, Rivera P, Perales A, Fayos J (1983). Structure and absolute configurations of *Dictyota* sp. diterpenes. Tetrahedron, 39: 3355-3357.
- Harden EA, Falshaw R, Carnacha SM, Kern ER, Prichard MN (2009). Virucidal activity of polysaccharide extracts from four algal species against herpes simplex virus. Antiviral Res., 83: 282-289.
- Kelecom A, Teixeira VL (1988). Dolastane diterpenes from the marine alga *Dictyota cervicornis*. Phytochemistry, 27: 2907-2909.
- Morfin F, Thouvenot D (2003). Herpes simplex virus resistance to antiviral drugs (review). J. Clin. Virol., 26: 29-37.
- Newman DJ, Cragg GM, Snader KM (2003). Natural product as sources of new drugs over the period 1981-2002. J. Nat. Prod., 66: 1022-1027.
- Pereira HS, Leão-Ferreira LR, Moussatché N, Teixeira VL, Cavalcanti DN, Costa LJ, Diaz R, Frugulhetti ICPP (2004). Antiviral activity of diterpenes isolated from the Brazilian marine alga *Dictyota menstrualis* against human immunodeficiency virus type 1 (HIV-1). Antiviral Res., 64: 69-76.

- Pereira HS, Leão-Ferreira LR, Moussatché N, Teixeira VL, Cavalcanti DN, Costa LJ, Diaz R, Frugulhetti ICPP (2005). Effects of diterpenes isolated from the Brazilian marine alga *Dictyota menstrualis* on HIV-1 reverse transcriptase. Planta Med., 71: 1019-1024.
- Reed B, Munch MA (1938). Simple method of estimating fifty per cent endpoints. Am. J. Virol., 27: 492–504.
- Teixeira VL,Tomassini T, Fleury BG, Kelecom A (1986a). Dolastane and secodolastane diterpenes from the brown alga *Dictyota cervicornis.* J. Nat. Prod., 49: 570-575.
- Teixeira VL, Tomassini T, Kelecom A (1986b). Cervicol a further secodolastane diterpene from the marine brown alga *Dictyota cervicornis* Kützing (Phaeophyceae Dictyotaceae). Bull. Soc. Chim. Belg., 95: 263-268.
- Vallim MA, De-Paula, JC, Pereira, RC, Teixeira VL (2005). The diterpenes from Dictyotaceae marine brown algae in the tropical Atlantic American region. Biochem. Syst. Ecol., 33: 1-16.
- Witvrouw M, De Clercq E (1997). Sulfated polysaccharides extracted from sea algae as potential antiviral drugs. Gen. Pharmac., 29: 497-511.
- Zanfardino A, Gavagnin M (2009). Diterpene content of the alga *Dictyota ciliolate* from a Moroccan lagoon. Phytochemistry Lett., 2: 211-215.